nature portfolio

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Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our <u>Editorial Policies</u> and the <u>Editorial Policy Checklist</u>.

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For	all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.
n/a	Confirmed
	\square The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
\boxtimes	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
\boxtimes	A description of all covariates tested
	🔀 A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
\boxtimes	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
\boxtimes	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
\boxtimes	\square Estimates of effect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated
	Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.

Software and code

Policy information about <u>availability of computer code</u>

Data collection No software was used.

Data analysis PatchMaster v1.4; GraphPad Prism9; MAFFT v7.304b; IQ-TREE v2.0.6; ModelFinder; PAUP* v4.0a;

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio guidelines for submitting code & software for further information.

Data

Policy information about <u>availability of data</u>

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our policy

The data that support the findings of this study are available within the article, Supplementary Information, or Source Data file. Sequences of Ta3a, Ta2a, Pc1a, Rm4a and Mri1a are available in Genbank: OW518818.1, Genbank: OW518839.1, UniProt: P41736, GenBank: MW317032 and GenBank: MN765042.1, respectively.

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	Not applicable			
	Not applicable			
ation on the app	roval of the study protocol must also be provided in the manuscript.			
	is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.			
	Behavioural & social sciences			
the document with	all sections, see <u>nature.com/documents/nr-reporting-summary-flat.pdf</u>			
nces st	udy design			
close on these	points even when the disclosure is negative.			
Sample size ca	Iculations were done with an alpha of 0.05 for 80% power where appropriate.			
No data were	excluded.			
All data is pres	ented with a minimum of 3 biological replicates. All replicates yielded similar results.			
For in vivo experiments, animals were allocated to treatments by complete randomisation with allocation concealment. Randomization was not performed in in vitro experiment with pairwise design such as electrophysiological recordings pre- and post-toxin exposure.				
Experiments involving pain behaviors, consisting of paw licks or flinches, were counted from video recordings by a blinded investigator (i.e. blinded to group allocation). Blinding was not relevant to other in vitro experiments presented as measurements were analyzed by applying the same automated criteria.				
	pecific materials, systems and methods about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, by your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.			
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Eukaryotic cell lines

Policy information about cell lines and Sex and Gender in Research

Cell line source(s) Human Embryonic Kidney (HEK) 293 cells (American Tissue Culture Collection, Manassas, VA, USA)

Stable HEK293-NaV1.6 (SB Drug Discovery, Glasgow, United Kingdom) Stable HEK293-NaV1.7 (SB Drug Discovery, Glasgow, United Kingdom)

Stable CHO-NaV1.8 (ChanTest, Ohio, USA)

F11 (Sigma Aldrich Australia, European Cell Culture Collection)

Authentication None of the cell lines used were authenticated

Mycoplasma contamination The cell lines were not tested for mycoplasma contamination

Commonly misidentified lines (See ICLAC register)

No commonly misidentified cell lines were used in this study

Animals and other research organisms

Policy information about <u>studies involving animals</u>; <u>ARRIVE guidelines</u> recommended for reporting animal research, and <u>Sex and Gender in Research</u>

Laboratory animals

1) 5–8 week old C57BL/6J mice (male) housed in groups of up to four per cage, maintained on a 12/12 h light-dark cycle (19-21 degrees celcius; 60-70% humidity), and fed standard rodent chow and water ad libitum.

2) Blowflies, Lucilia caesar; adult (1-4 d post-emergence; average mass 19 mg)

Wild animals

Adult female T. africanum (worker caste; age unspecified) used for venom collection were collected from in Cameroon. They were collected, transported and maintained (briefly) in a plastic container. Specimen were dissected for collection of venom after which

remaining tissue was discarded.

Reporting on sex For experiments involving mice and mouse tissues, the sex of mice is reported and no sex- and gender-based analyses have been

performed. The applicability of the findings of this study are independent of sex or gender.

Field-collected samples This study did not involve samples collected in the field.

Ethics oversight Experiments involving animals were approved by The University of Queensland Animal Ethics Committee (UQ AEC approval numbers PHARM/526/18 and 2021/AE000448). Experiments involving use of mouse tissue were approved by the UQ AEC (approval TRI/

IMB/093/17).

Note that full information on the approval of the study protocol must also be provided in the manuscript. $\frac{1}{2} \int_{\mathbb{R}^{n}} \left(\frac{1}{2} \int_{\mathbb{R}^{$